Diagnosis of otitis externa, media and interna in rabbits

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Aural disease in rabbits is becoming increasingly diagnosed in clinical veterinary practice and is often associated with chronic upper respiratory disease.

Figure 1. Head tilt is a common presentation in rabbits.

One of the common signs of otitis media and interna are vestibular disease and this is often erroneously considered to be due to *Encephalitozoon cuniculi*. Head tilt, nystagmus and torticollis are common presenting clinical signs in rabbits (Figure 1).

The most important differentials for this condition are *E cuniculi* causing central vestibular disease or *Pasteurella multocida* causing otitis media, which leads to otitis interna, and the clinical signs become evident due to pressure and disruption of the vestibular apparatus.

Presumptive therapy usually centres on treating these two different conditions. Many cases are
treated with oral fenbendazole for the *E cuniculi* and the patient may be given antibiotics such as enrofloxacin. Nonsteroidals or, in some cases, steroids may be given. Supportive care measures are vital as severe cases are unable to feed and gastrointestinal stasis is inevitable. Euthanasia should be considered. Severe cases may require sedation to reduce the severity of any spiralling.

Vestibular disease is reported as a frequent manifestation of *E cuniculi* infection. However, to lead to such specific signs the inflammation created by the parasite must be in a single focus. This is unlikely and greater effort should be made to confirm that *E cuniculi* is, in fact, causing the clinical signs, primarily by ruling out the other most important cause – otitis media and interna. If both are negative, more unusual diagnoses may be relevant.

**Figure 2.** The central ear artery can be used for blood gas analysis.

In peripheral vestibular disease, the vestibular nerve (cranial nerve VIII) and the labyrinth of the inner ear are affected, usually as an extension of otitis media. This could be due to simple pressure on the oval and round windows, inflammation or infection. These cases have a horizontal nystagmus, which can be positional or evoked.

Central vestibular disease is when the lesions are in the brainstem. These present with vertical nystagmus. This is more likely with encephalitis. Other nervous defects, such as paresis or reduced mentation, may be present if the disease is extensive or multifocal, which, of course, does not occur with otitis media and interna.

In a UK study, 52 per cent of rabbits had IgG antibodies to *E cuniculi*, indicating these rabbits had been exposed to this agent (Keeble, 2006). This result often leads to treatment of the rabbit with fenbendazole with no further diagnostics being performed to eliminate other causes. IgG antibodies elevate four weeks after infection, peak at nine weeks and then slowly decline. Paired titres may actually yield more useful information and are generally taken a month apart. An increased second titre suggests the infection is active. A negative IgG antibody does not rule out *E cuniculi*, as the
infection may be early or latent.

IgM titres have become available and elevate quickly after infection and decrease to a low level by a month after infection. A high titre indicates recent active infection or a recent reactivation of a latent infection. This enables the clinician to differentiate between exposure and active (new or reactivated) infections.

![Vertical acoustic meatus](image)

**Figure 3.** The bulla has thick bone laterally and thinner bone ventrally.

Generally, IgG and IgM titres are run concurrently and compared. Approximately a third (32.8 per cent) of rabbits have IgM antibodies and this increases (to 54.4 per cent) if the rabbits have neurological disease (Jecklova, 2010). Given this, a large number of neurological rabbits may not have active *E cuniculi* infection, but it takes 70 to 100 days after infection for lesions to be identified in the CNS. High IgM titres would, therefore, not be expected in neurological cases unless a latent infection had been reactivated at the same time.

Antibody titres alone do not confirm the clinical diagnosis. One paper demonstrated that a cerebrospinal fluid (CSF) tap proved useful as there were elevated white cell counts (lymphomonocytic pleocytosis) if encephalitis was present (Jass, 2008). This could be due to any CNS inflammation and ruling out other CNS diseases such as toxoplasmosis is important. Laparoscopic biopsies may aid diagnosis, yielding signs of lymphocytic inflammation. Although urine PCR has become available, shedding only occurs for five weeks post-infection. Other less common differentials for nervous diseases include toxoplasmosis, herpes simplex, listeriosis or other degenerative CNS lesions (Keeble, 2011).

**Aural anatomy in rabbits**

Knowledge and experience of treating otitis in dogs and cats is an important prerequisite for
treated otitis in rabbits. However, rabbit aural anatomy, disease pathogenesis and treatment are different.

![Angular process](image)

**Figure 4.** The angular process of the mandible is a useful landmark.

The external ear is made of a pinna and a vertical canal. The pinna has a central artery with many veins laterally, any of which can be catheterised if large enough, but most usually the caudal vein is utilised. The central ear artery is usually avoided for catheterisation, but can easily be used for blood gas analysis (**Figure 2**). Neovascularisation (to bypass previous sites of catheterisation) is very common. Pinna size is variable. Many breeds have an upright pinna, but lop breeds have a pinna that is folded downwards. Rabbits do not have separate vertical and horizontal canals, but have multiple cartilaginous plates making up the single “vertical” ear canal that extends dorsally from the bony acoustic meatus. This leads to the tympanic membrane at the ventral aspect of the meatus at the point of entry to the bulla. The bulla consists of very thick bone laterally and thinner bone ventrally and projects ventral and lateral to the base of the acoustic duct (**Figure 3**). The bulla is 5mm deep, 7.5mm high and 11mm long (Chow, 2011; Mayer, 2011; Popesko et al, 1992).

Lop breeds are predisposed to otitis due to their altered anatomy (Chow, 2011; Capello, 2004). In these breeds they have a fold in the vertical canal between these plates and the canal is stenotic.

Rabbits have a much wider mandible than dogs and cats at its caudal extent. This extends ventrally in a semicircular fashion. This reduces the ability to palpate the bulla in a conscious rabbit. The angular process of the mandible protrudes caudally just below the entrance point to the bulla.
and provides a useful landmark (Figure 4).

**Clinical signs associated with ear disease in rabbits**

**Otitis externa**

Clinical signs of otitis externa include scratching at the base of the ear, head shaking, pain, lethargy or anorexia. Primary bacterial infection or dermatopathies leading to otitis externa are rare. Rabbit wax is thick and appears similar to pus. However, wax is typically yellow or beige in colour (Figure 5). Pus is typically a white or creamy colour. Cytology can be used to confirm if there are large numbers of white cells or bacteria if otitis externa is present. *Psoroptes cuniculi* is a common parasitic problem and the diagnosis is essentially clinical (Figure 6). However, skin scrapes can be taken to confirm infection.

**Otitis media**

Otitis media cases are often clinically silent, but may be identified on imaging for another reason. These cases often present with lethargy, inappetence, pain, pruritus associated with the base of the ear, a head wobble, or a painful swelling at the base of the ear (Figure 7). Owners may report hearing deficits, but this is unusual.

**Figure 5.** Rabbit wax is thick and similar to pus, but a beige or yellow colour.
Figure 6. *Psoroptes cuniculi* is a common parasitic problem.
**Figure 7.** A swelling at the base of the ear can indicate otitis media.

Facial paralysis or spasticity may occur on the side of the lesion (**Figure 8**) due to extension of the pathology to involve the nerve around the lateral side of the bulla and ventral to the ear canal. Infection can extend to the external ear canal if the tympanic membrane has been ruptured. Alternatively, infection can progress from otitis external to otitis media. Unilateral and bilateral disease is seen. Otitis media may extend to otitis interna.

**Otitis interna**

![Figure 8. Left side facial paralysis on the side of the lesion in a case of otitis media.](image)

Otitis interna is usually present with otitis externa. It can occur due to pressure, inflammation or infection. Head tilt, nystagmus, ataxia and circling are common signs. Facial nerve spasticity, paresis or deficits, may be present as the facial nerve exits ventral to the vestibulocochlear nerve in the internal acoustic meatus. Drooping of the upper lip, eyelid and ear (Horner’s syndrome) may be present and mimic facial paralysis.

**Pathogenesis of otitis externa**

Ceruminous gland hyperplasia does not occur, even with severe otitis externa. In cats and dogs this hyperplasia leads to narrowing of the ear canal. These changes are primarily in the vertical canal and so a lateral wall resection may be indicated in these species.

Cases of otitis externa in rabbits can be an extension of otitis media via the ruptured tympanic
membrane, resulting in infection at the base of the vertical canal.

In severe cases, the local tissues can become involved and degeneration of the ear canal leads to a soft tissue swelling at the base of the ear canal. The infection rarely advances up the canal and it does not lead to marked histological changes of the dorsal section of the vertical canal.

**Figure 9.** Sebaceous ductal ectasia.

Mild cases have minimal histopathological changes limited to subtle and nonspecific changes, such as hyperkeratosis or mild sebaceous ductal ectasia in the vertical canal (**Figure 9**). It is difficult to distinguish between these two disease processes clinically, but otitis media extending to otitis externa is by far the most frequent presentation.

**Pathogenesis of otitis media or interna**

Those cases with clinical signs of upper respiratory tract disease or those that are positive for *Pasteurella multocida* are highly likely to suffer from this condition due to infection ascending via the Eustachian tubes.

In one rabbit study, *P. multocida* was isolated in most clinical and subclinical cases of otitis media and interna. However, isolation of *P. multocida* does not equate to disease and up to 95 per cent of rabbits may be positive (Deeb et al, 1990; Snyder et al, 1973; Smith and Webster, 1925).

In dogs, culture results yielded from tympanic bullae pre and post-flushing at surgery demonstrated a 33 per cent reduction in isolates, but 70 per cent of these isolates were different to those isolated preflushing and 84 per cent of ears had different sensitivity patterns (Hettlich et al, 2005), therefore, isolation of a specific pathogen in rabbits may be misleading.
Up to 30 per cent of rabbits with respiratory pasteurellosis in one study had subclinical otitis media and evaluation of the rabbit for underlying respiratory disease is vital. It is also possible for a rabbit to have a positive culture from the bulla, but be negative on nasal swabs. Other agents cultured from otitis media cases include *Bordetella bronchiseptica*, *Staphylococcus*, *Escherichia coli* and *Pseudomonas aeruginosa*.

### Diagnosis

**Table 1. Assessment of cranial nerve function in rabbits.**

<table>
<thead>
<tr>
<th>Cranial nerve</th>
<th>Function test</th>
</tr>
</thead>
<tbody>
<tr>
<td>CN I (Olfactory)</td>
<td>Response to noxious smelling substance.</td>
</tr>
<tr>
<td>CN II (Optic)</td>
<td>Menace response.</td>
</tr>
<tr>
<td>CN II (Optic)</td>
<td>Pupillary light reflex.</td>
</tr>
<tr>
<td>CN II (Optic)</td>
<td>Visual placing reflex.</td>
</tr>
<tr>
<td>CN III (Oculomotor)</td>
<td>Observe eye position and movements at rest and on movement of head (vestibulo-ocular reflex).</td>
</tr>
<tr>
<td>CN IV (Trochlear)</td>
<td>As for CN III.</td>
</tr>
<tr>
<td>CN V (Trigeminal)</td>
<td>Size and symmetry of masticatory muscles. Test resistance to jaw opening.</td>
</tr>
<tr>
<td>Motor function</td>
<td></td>
</tr>
<tr>
<td>CN V (Trigeminal)</td>
<td>Corneal reflex, Palpebral reflex, Facial skin pinching and observation of facial twitch.</td>
</tr>
<tr>
<td>Sensory function</td>
<td></td>
</tr>
<tr>
<td>CN VI (Abducent)</td>
<td>As for CN III.</td>
</tr>
<tr>
<td>CN VII (Facial)</td>
<td>Observe facial symmetry, blinking and nostril movement. Palpebral reflex, Corneal reflex, Menace response, Facial pinching.</td>
</tr>
<tr>
<td>CN VIII (Vestibulocochlear)</td>
<td>Response to sudden noise, Vestibulo-ocular reflex (nystagmus induced by head movement).</td>
</tr>
<tr>
<td>CN IX (Glossopharyngeal)</td>
<td>Observation of swallowing, Gag reflex.</td>
</tr>
<tr>
<td>CN X (Vagus)</td>
<td>Oculocardiac reflex.</td>
</tr>
<tr>
<td>CN XII (Hypoglossal)</td>
<td>Inspection of tongue, Observation of licking.</td>
</tr>
</tbody>
</table>

**Table 1.** Assessment of cranial nerve function in rabbits.

Clinical examination may yield clinical signs consistent with otitis externa, with distortion of the base of the vertical canal or purulent material being present. A neurological examination should be performed, particularly assessing the function of cranial VII, VIII and IX nerves (**Table 1**).

The facial nerve (VII) relays sensory information from, and provides motor activity to, the mandible, maxilla and eyelids. The rabbit should be able to open its mouth, blink and its nose should twitch. A small needle can be used to elicit a twitch of the muscles of the face. Head shaking or moving
away from this indicates a central component. The menace and corneal responses are also
dependent on facial nerve function.

The vestibulocochlear nerve (VIII) is evaluated by a response to a sudden loud noise or rattling a
food bag. Many owners may be able to provide some useful clinical history, but if the disease is
unilateral it can be difficult to quantify. The nerve is responsible for balance, and nystagmus may
be induced by postural changes. Placing and balancing tests can also be used.

The glossopharyngeal nerve (IX) is best assessed by tongue movement and taste. Enrofloxacin or
metronidazole can be dropped on the tongue, which should elicit a response.

Otoscopic examination (Figure 10) or endoscopic examination of the ear canal under anaesthesia
(Figure 11) can help confirm the extent of the infection. For endoscopy, the rabbit can be placed in
lateral or ventral recumbancy. Saline insufflation can be used to aid visualisation of the ear canal. It
can be difficult to identify the tympanic bulla if there is a large amount of material in the canal and
difficult to confirm rupture. Removal of material with cotton buds, flushing or grasping with
endoscopic forceps (Figure 12) may help, but runs the risk of mucosal trauma, leading to local
bleeding into the canal hindering visualisation further. However, endoscopic biopsies of tissue or
exudate deep within the canal can be submitted for culture and histopathology. As the biopsy
instrument is sterile until it is passed out the end of the endoscopic biopsy sheath this will yield
more reliable results than a culture swab passed down the ear canal, which may pick up more
superficial contaminants.

Positive contrast canalography is also a technique to consider. Contrast material may pass into the
bulla and confirm a ruptured tympanic membrane. If otitis media without externa is suspected, the
tympanic membrane can be evaluated endoscopically and an injection/ aspiration needle used to
obtain samples from within the bulla if fluid or purulent material is identified behind the tympanic
membrane that may also lead to a bulging of the membrane into the canal.

A myringotomy (opening the tympanic membrane) will allow for this material to pass into the
external ear canal (easing pressure and maybe reducing the risk of otitis interna), but a surgical
approach to the otitis media is preferred.
**Figure 10.** Otoscopic examination of the ear canal.

**Figure 11.** Endoscopic examination of the ear canal should be made with the patient anaesthetised.
Figure 12. Removal of material runs the risk of mucosal trauma leading to bleeding into the ear canal.

Radiography is commonly performed under anaesthesia or sedation. The dorsoventral view is reported as being the most useful (Figure 13) although lateral, oblique views and open mouth cranial caudal views can also be taken. Oblique views should be between 30 and 70 degrees to visualise the bullae separately. Increased sclerosis of the bone or radio-opacity of the bulla are often reported to be the identifying signs of otitis media. Radiography is a poor diagnostic technique in dogs and many cases of otitis media in rabbits may well be missed. This generally results in a tentative diagnosis at best and many cases are treated medically as a result.

The accumulation of fluid or pus within the tympanic bulla is an important diagnostic indicator to confirm otitis media. CT has been confirmed as the most reliable method of diagnosis in dogs, although ultrasound also shows promise (Rohleder, 2006; Dickie, 2003). Radiography yields
accurate results only in 56 per cent of cases. In a cadaveric study in rabbits, ultrasound has been found to be useful in achieving a diagnosis (King, 2007). CT is the best imaging modality for evaluation of the skull. High-resolution images can be obtained with a sedated rabbit in a short timeframe (Figure 14). Contrast is not required when evaluating the bullae. Interpretation is simple since otitis media is clearly identified due to increased soft tissue density being present where the bulla should be gasfilled (Figure 15). Otitis externa is more challenging because both pus and wax will fill the vertical canal and have a similar appearance on CT. Material can be seen to arise from the external ear canal entering the bulla, confirming tympanic membrane rupture (and suggests extension of otitis externa to media), while in others (much more commonly) the bullae is completely filled with a soft tissue density. A third group may have a soft tissue density at the base of the bulla that has not progressed to filling the whole canal. At our clinic, radiographic assessment is no longer used for identifying otitis media as these changes are impossible to identify radiographically.

Diagnosing otitis media is simple with CT, but linking its clinical significance to the clinical signs (if present) is harder. Many rabbits will have subclinical infections (identified due to CT screening for respiratory or dental disease, for example). It is unclear at what point clinical disease will, if ever, become evident, but the current suggestion is that a soft tissue density in the bulla is pathological. In one postmortem study in laboratory rabbits 32 per cent of the tympanic bullae were found to have purulent contents (otitis media) that was clinically silent at the time of death (Snyder, 1973).
Figure 13. The dorsoventral view is usually the most useful.
Figure 14. Computed tomography is the best imaging modality for the skull.
It is impossible to quantify the level of chronic pain present with this condition. The author’s opinion is any cases of otitis media identified on CT examination should be treated, with a view to obtaining a complete resolution. It is unclear if neurological signs will improve or resolve with medical or surgical treatment of otitis media. However, if the otitis media is generated by pressure being applied to the oval or round windows then alleviating the pressure by surgical removal may provide the best option for resolution. Quality of life should be taken into account when considering surgery on those rabbits with marked neurological signs.

- Please note drugs mentioned in this article are used under the cascade.
References